

Urinary excretion of lipid mediators in response to repeated eucapnic voluntary hyperpnea in asthmatics

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42 **Abstract**

43 Exercise-induced bronchoconstriction displays refractoriness manifested as a decreased
44 response to repeated exercise challenge within hours. The refractoriness may be attenuated
45 by inhibition of the biosynthesis of prostaglandins (PG). The aim of the study was to
46 determine which PGs and other lipid mediators are excreted during the refractory period.

47 First, 16 subjects with mild stable asthma performed two repeated 4 min challenges
48 with eucapnic voluntary hyperpnea (EVH) 1 and 3 hours apart. There was a similar degree
49 of refractoriness in both protocols (about 15% protection). The one hour interval was too
50 short to study mediator excretion because the urinary levels did not return to baseline before
51 the second challenge. With the 3 hour protocol, there was increased urinary excretion of
52 cysteinyl-leukotrienes (CysLTs) and metabolites of the mast cell product PGD₂ after both
53 challenges.

54 Next, another 8 subjects performed two 6 min challenges with EVH 3 hours apart
55 which produced a greater bronchoconstrictor response than the 4 min protocol(30.0±5.4% vs
56 17.7±1.5%; p=0.0029) and a greater degree of refractoriness (about 30%) Analysis by
57 UPLC-MS/MS confirmed excretion of the bronchoconstrictor CysLTs and PGD₂ during both
58 challenges. In addition, there was increased excretion of the bronchoprotective PGE₂, and
59 also of the main metabolite of PGI₂.

60 This is the first report of excretion of PGE₂ and PGI₂ during the refractory period to
61 EVH challenge, suggesting that they may mediate the refractoriness. Maintained excretion of
62 PGD₂ and LTE₄ following the repeat challenge argues against mast cell mediator depletion as
63 the mechanism of refractoriness.

64 **Word count: 246**

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67 **Introduction**

68 Exercise is a common trigger for an attack of asthma and a majority of asthmatic subjects
69 experience bronchoconstriction after exercise.(4) The narrowing of the airways is thought to
70 be caused by dehydration and a transient hyperosmolarity of the airway surface which causes
71 release of bronchoconstricting mediators from mast cells and other cells in the airways.(2) In
72 support of hyperosmolarity being the mechanism triggering exercise-induced
73 bronchoconstriction (EIB), similar airway responses are seen following inhalation of
74 mannitol,(8, 22) and eucapnic voluntary hyperpnea (EVH)(5, 32). These challenges are
75 therefore being used as surrogates for exercise to diagnose potential for EIB.(17) The
76 bronchoconstricting mediators released into the airways are rapidly removed by the
77 circulation and are excreted into the urine.(20, 27) Using enzyme immuno assay (EIA)
78 methodology, we have previously demonstrated increased urinary excretion of cysteinyl
79 leukotrienes (CysLTs) and the prostaglandin (PG) D₂ metabolites following exercise(28),
80 EVH(18, 19), and mannitol(22), respectively. Whereas CysLTs may be biosynthesised in
81 many inflammatory cells, PGD₂ is almost exclusively produced by the mast cells and its
82 release therefore provides objective evidence of mast cell activation. Previous knowledge
83 about mediator excretion into the urine following EIB has however been restricted to those
84 two lipid mediators.

85 Here we report on an extended spectrum of mediators using a newly developed platform
86 for mass spectrometry enabling us to simultaneously study the urinary excretion of
87 metabolites of CysLTs, PGD₂, PGE₂, PGF_{2α}, thromboxane A₂ (TXA₂), as well as multiple
88 isoprostane species.(6) The methodology was applied to establish the profile of lipid
89 mediators generated in response to repeated EVH challenges. When challenge by exercise, or
90 with EVH/mannitol, is repeated within 4 hours, a decrease in the bronchoconstrictor response
91 is observed.(11) This decreased responsiveness to repeated challenges is called refractoriness

92 and its duration the refractory period. The occurrence and the degree of refractoriness
93 decrease continuously with increasing time between the challenges.(11) The mechanisms
94 behind this protective response remain unclear. We hypothesise that identification of
95 endogenous molecules that mediate refractoriness may help to define new targets for
96 treatment of airway obstruction.

97 We have previously observed that the subjects who were most refractory to repeated
98 challenge with mannitol had the highest levels of CysLT and the PGD₂ metabolite 11β-
99 Prostaglandin (PG) F_{2α} during the refractory period.(22) This finding suggested a decreased
100 responsiveness to the released mediators at the level of the airway smooth muscle as one
101 possible mechanism in refractoriness.(21, 22) Because non-steroidal anti-inflammatory drugs
102 (NSAIDs) almost completely abolish refractoriness (26), it has been proposed that PGE₂ may
103 be of importance for the development of refractoriness.(24, 26, 36) However, there is to date
104 no direct evidence for release of PGE₂ *in vivo* during the refractory period.

105 Because the occurrence of refractoriness depends upon the time between the challenges
106 and because the urinary excretion of mediators also has time-dependent kinetics, our study
107 first defined the optimal conditions for this combined study of the bronchoconstrictor
108 response and the urinary excretion of alleged mediators of bronchoconstriction and
109 refractoriness. After the establishment of a suitable experimental design, it was possible to
110 provide the first evidence of increased excretion of PGE₂ following EVH challenge. The
111 study also discovered increased urinary excretion of metabolites of PGI₂ during the refractory
112 period, possibly adding yet another potential endogenous protective factor to consider.

113

114 **Materials and Methods**

115 *Study design*

116 The urinary excretion of mediators is delayed compared to the airway response necessitating
117 us to determine the optimal interval between challenges to detect refractoriness while still
118 being able to study the urinary mediator excretion. Therefore, we performed an initial study
119 (study 1) to compare two different intervals between challenges. In this first study 16
120 asthmatics were recruited to perform repeated 4 min EVH challenge either 1 or 3 hours apart,
121 in a randomised cross-over design.(figure 1)

122 During screening the subjects underwent a physical examination, skin prick test, and
123 spirometry. A 4 min EVH challenge was performed using a slight modification of a protocol
124 published by Smith et al.(35) Subjects who met the inclusion criteria of a maximum fall in
125 $FEV_1 \geq 10\%$ were included in the study. On the two study days, following baseline
126 spirometry, repeated challenge with 4 minutes of EVH (challenge I and challenge II) was
127 performed. Urine samples were collected 30 minutes before, immediately before the start of
128 the first challenge and then hourly until 240 minutes after the first challenge. Lung function
129 was monitored repeatedly. In order to be able to calculate a percentage protection, data were
130 analysed per protocol excluding the subjects who did not achieve a 10% fall in FEV_1 on the
131 first challenge on a particular study day. For the one hour protocol, 5 of the 16 and for the 3
132 hours protocol 1 of 16 subjects did not achieve a fall in $FEV_1 \geq 10\%$ following challenge I,

133 Based upon this initial range-finding study, study 2 was designed. Now, a 6 minute EVH
134 challenge was performed and only subjects who had a maximum fall in $FEV_1 \geq 15\%$ were
135 included. Nine subjects met the inclusion criteria, but one subject was excluded from analysis
136 because of asthma deteriorating between the screening visit and the study day, indicating that
137 she did not meet the inclusion criteria of having stable mild asthma. Thus 8 subjects were
138 eligible for further analysis. The screening day was performed in the same way as for study 1

139 with the exception of 6 min EVH instead of 4 min EVH. During the study day the subjects
140 performed repeated challenge with 6 minutes of EVH 3 hours apart. Urine samples were
141 collected 30 minutes before, immediately before the start of the first challenge and then every
142 hour until 300 minutes after the first challenge.

143

144 *Subjects*

145 Non-smoking subjects with mild and stable asthma were eligible for participation. Asthma
146 was defined by at least one of three criteria; response to asthma treatment, episodic wheezing
147 and variation in lung function over short periods of time. To be included the subjects had to
148 display baseline FEV₁ ≥70% of predicted value. Study subjects only used asthma medications
149 as needed and were allowed to have used short acting β₂-agonist only during the month before
150 the study. Exclusion criteria included respiratory tract infection within the last six weeks
151 before inclusion. Subject characteristics are presented in table 1.

152 All included subjects gave their written informed consent and the study was approved by
153 the local ethics committee (Karolinska Institutet regional ethics committee Dnr 03-127, Ethics
154 board Stockholm 2012/1277-32).

155

156 *EVH*

157 Hyperpnea with dry, room temperature air containing 5% carbon dioxide was performed
158 through a low-resistance, one-way valve in the sitting position (Ailos Asthma Test[®],
159 Karlstad, Sweden) (5, 32). The target ventilation was 35 x FEV₁ x 0.75 (L/min) and was
160 maintained for 4 or 6 minutes.

161

162 *Lung function*

163 Lung function (FVC and FEV₁) was measured according to the American Thoracic Society
164 criteria, using a wedge spirometer (Vitalograph[®], Buckingham, UK). FEV₁ was measured in
165 duplicate before EVH, immediately after, at 2, 5, 10, and then every 10 minutes during 1 hour
166 following each challenge. On each occasion the highest value of two FEV₁ measurements was
167 registered. The fall in FEV₁ was calculated in percent of the pre-challenge value.

168

169

170 *Skin prick test*

171 Skin prick test was performed during screening using the following allergens; birch, timothy,
172 mugwort, dog, cat, horse, dermatophagoides pteronyssinus, farinae, *cladosporium* and
173 *alternaria* (Soluprick SQ, ALK, Denmark). A positive response was defined as a measurable
174 wheal of ≥ 3 mm in the absence of any equivalent reaction in the control test.

175

176 *Urinary mediators*

177 After collection, urine samples were stored at -70°C until analysis. All urine samples were
178 analysed for creatinine using the modified Jaffe colorimetric method.(22) LTE₄, 11 β -PGF_{2 α} ,
179 8-isoprostane-PGF_{2 α} , 6-keto-PGF_{1 α} , PGE₂, TXB₂ and tetranor-PGDM were analysed using
180 enzyme immunoassay kits commercially available (Cayman Chemical, Ann Arbor, Michigan)
181 as previously described.(22) In addition, the urine samples from study 2 were analysed using
182 ultra-performance liquid chromatography triple quadrupole mass spectrometry (UPLC-
183 MS/MS) as described by Balgoma et al.(6), with the exception that prostacyclin metabolites
184 not were detected due to a technical error. All levels of mediators were corrected for dilution
185 using creatinine and expressed as ng/mmol creatinine.

186

187 *Statistical analysis*

188 All data are presented as mean value \pm standard error of the mean (SEM), unless otherwise
189 stated. Statistical significance was determined using paired t-test to identify differences in the
190 maximum fall in lung function and to compare baseline with peak mediator excretion. The
191 Wilcoxon signed rank test was performed for data that were not normally distributed.
192 Correlations were calculated using Pearson product moment correlation. Significance was
193 defined as the commonly accepted level of p-value of <0.05 . All statistical analyses were
194 performed using GraphPad Prism version 5.00 for Windows, GraphPad Software, San Diego
195 California USA, www.graphpad.com.
196

197 **Results**

198 Study 1 - Defining optimal time between challenges

199 Subject characteristics of the 16 subjects included in this first part of the study are presented
200 in Table 1. Skin prick test was positive in 13 of the subjects. There were no significant
201 differences in pre-challenge lung function (FVC and FEV₁) or exhaled NO levels on the
202 different study days (data not shown).

203 *Airway response* - For the one-hour protocol (n =11) the mean maximal decrease in FEV₁
204 was 19.6±2.4 % after EVH challenge I and 15.4±1.2 % following challenge II one hour later
205 (p = 0.06). For the three-hour protocol (n = 15) the mean maximal decrease in FEV₁ was
206 19.5±2.1 % after challenge I and 16.3±2.5 % after challenge II three hours later (p = 0.02)
207 (table 1). These results correspond, to an attenuation of 15 % when challenge II was
208 performed after one hour, and 19 % attenuation when challenge II was done three hours after
209 challenge I. No correlation was found between % fall following challenge I and the protection
210 afforded for neither of the 1 hour nor the 3 hour study day. Comparing the degree of
211 protection between the 1 hour and 3 hour protocols for the 11 subjects who had data from
212 both study days we found no differences; the mean protection was 15±9% and 15±8%
213 respectively.

214 *Urinary CysLT and PGD₂ excretion* - with the 1 hour protocol (n=11) there were no
215 significant differences in the excretion (ng/mmol creatinine [±SEM]) of LTE₄ or 11β-PGF_{2α}
216 after either of the challenges (figure 2A and 2C).

217 In contrast, for the 3 hour protocol (n=15) 11β-PGF_{2α} increased significantly both after
218 challenge I (29±4 vs 24±4 ng/mmol creatinine, p=0.0054) and after the challenge II (28±3 vs
219 24±3 ng/mmol creatinine, p=0.0209) (figure 2B). Also, there was an increase of LTE₄ after
220 challenge I (40±3 vs 32.5±3 ng/mmol creatinine, p=0.0011) and a strong tendency for an
221 increase after challenge II (39±4 vs 33±3 ng/mmol creatinine, p=0.0582) (figure 2D).

222 *Repeatability of the EVH challenge* - The agreement between challenges was in general
223 good, however in a few patients the standard deviation was sometimes large, with differences
224 >20% when comparing the maximum percentage fall in FEV₁ following challenge I. Bland-
225 Altman plots for demonstration of the variability revealed that the larger the fall in FEV₁ the
226 greater the variability becomes. (7) There were however no significant differences between
227 the screening day and the study days (table 1).

228

229 Study 2 – extended analysis of urinary mediator excretion

230 From the results of study 1 it was obvious that the one hour interval between the challenges
231 was too short and unsuitable because the mediator levels do not return to baseline within this
232 time frame. Also, since the timing of the peak excretion of mediators after a challenge differ
233 between subjects it is not satisfactory with only one sample following challenge. Therefore
234 another 8 subjects were recruited to perform repeated EVH 3 hours apart. In order to increase
235 the probability to catch the peak excretion following the second challenge, a sampling of urine
236 was added at 300 minutes after the first challenge. Also, to try to enhance the response, we
237 chose to use 6 minutes of EVH instead of 4 minutes and included only subjects with a fall in
238 FEV₁ ≥15% at screening. Subject characteristics are presented in table 1. Skin prick test was
239 positive in in 7 of the subjects

240 *Airway response* - The maximum fall in FEV₁ following challenge I was 30.0 ±5.4% and
241 21.2 ±4.5 % following challenge II, p=0.0076 (Table 1). The mean degree of protection was
242 30 % and all subjects showed some degree of protection, with the range being 2% to 70% ()
243 Thus, overall the bronchoconstriction induced in study 2 was greater than in study 1
244 (30.0±5.4% vs 19.5±2.1%. p=0.0029), indicating the 6 min EVH to be more powerful than the
245 4 min.

246 Extended urinary mediator analysis using both UPLC-MS/MS and EIA- The results of the
247 extended analysis are presented in table 2 (UPLC-MS/MS), in table 3 (EIA) and in figure 3A-
248 J (UPLC-MS/MS and EIA). Of the 30 metabolites in the platform, 13 were found at
249 measureable levels. The most abundant mediator was tetranor-PGEM (~3500 ng/mmol
250 creatinine) followed by the main isoprostane metabolite 8,12-iPF_{2α}-VI (~500 ng/mmol
251 creatinine). The tetranor metabolite of PGD₂ displayed levels around 200-250 ng/mmol
252 creatinine, whereas the levels of PGF_{2α} were around 100 ng/mmol creatinine. Metabolites of
253 thromboxane were slightly less abundant and CysLTs were found in the lowest concentrations
254 of all measured compounds. In general, the levels detected in EIA were higher compared to
255 the levels found using UPLC-MS/MS. There was however a good correlation between the
256 EIA and UPLC-MS/MS results.

257 PGD₂ metabolites – Measuring the peak excretion of 11β-PGF_{2α} using EIA the increase
258 from baseline did not reach significance following challenge I. Following challenge II there
259 was however a significant increase from baseline (table 3, figure 3A). The levels of 11β-
260 PGF_{2α} were back to baseline before challenge II. Using UPLC-MS/MS, while the early
261 metabolite 11β-PGF_{2α} was undetectable, its metabolite 2,3-dinor-PGF_{2α} was found in similar
262 concentrations as the 11β-PGF_{2α} values indicated by the EIA. The 2,3-dinor-PGF_{2α} increased
263 to an equal magnitude after both challenges indicating a similar level of mast cell activation
264 (table 2, figure 3B). The levels of these two early PGD₂ metabolites returned to baseline
265 before challenge II was initiated.

266 The EIA for tetranor-PGDM, a later and more abundant metabolite of PGD₂, showed
267 increased levels following challenge I, but failed to reach significance following challenge II
268 (table 3, figure 3C). Conversely, with UPLC-MS/MS, the increase of tetranor-PGDM failed
269 to reach significance following challenge I, but increased significantly following challenge II
270 (figure 3D).

271 Cysteinyl leukotrienes – Using EIA, for LTE₄ there was an increase from baseline
272 following the challenge I, but not following challenge II (table 3, figure 3E). The levels of
273 LTE₄ were still somewhat elevated from baseline before challenge II making interpretations
274 of excretion following challenge II more difficult. In UPLC-MS/MS, LTE₄ was the only
275 CysLT that could be detected (table 2). Most of the subjects displayed increased levels
276 following challenge I with a strong tendency for the whole group although not statistically
277 different. The levels returned to baseline before challenge II, and significantly increased
278 levels were seen following challenge II (figure 3F).

279 Thromboxanes and Isoprostanes – TXB₂ and its metabolites, the isoprostanes and PGF_{2α}
280 failed to display consistent increases irrespective of whether analysed by EIA or UPLC-
281 MS/MS. (table 2 and table 3).

282 Prostaglandin E₂ – The EIA for PGE₂ showed increased concentrations following
283 challenge I, but not following challenge II (table 3, figure 3G). In UPLC-MS/MS the levels of
284 PGE₂ were significantly increased following both of the challenges, with no differences
285 between the two peaks (table 2, figure 3H). Tetranor-PGEM increased significantly following
286 challenge I whereas the increase following challenge II failed to reach significance (table 2,
287 figure 3I). There were no differences in the levels of urinary PGE₂ between male and female
288 subjects, whereas the levels of PGEM were significantly higher in male subjects (The females
289 are #22, 23, and 24 in figure 3I). The peak levels of PGE₂ or PGEM after the first or second
290 challenge did not correlate with the degree of refractoriness.

291 Prostacyclin – Using the EIA clear increases were seen of the prostacyclin metabolite 6-
292 keto-PGF_{1α} following both the challenges (table 3, figure 3J). This metabolite was not
293 included in the UPLC-MS/MS. The peak levels of PGI₂ after the first or second challenge did
294 not correlate with the degree of refractoriness.

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321 **Discussion**

322 In this study we report on extended analysis of mediator excretion during the refractory period
323 following repeated EVH challenge. For the first time in this setting for exercise-induced
324 bronchoconstriction, we could demonstrate increased urinary excretion of metabolites of
325 PGE₂ and PGI₂ following EVH. In contrast, the levels of metabolites of thromboxane and
326 isoprostanes remained unchanged, indicating specificity in the excretion of eicosanoids.
327 Using the mass spectrometry platform we also documented increased levels of CysLTs, and
328 metabolites of PGD₂ following the EVH challenge, thus replicating and validating by mass
329 spectrometry our previous findings using EIA only.(18, 19) The increased urinary excretion
330 of those two bronchoconstrictive mediators following both of the two repeated EVH
331 challenges are similar to our previous findings of mediator excretion following repeated
332 mannitol inhalation challenge.(22) Taken together, the main findings support that excretion
333 of mast cell mediators is maintained also during the second challenge, and that there is
334 significant excretion of two lipid mediators with bronchoprotective properties during the
335 refractory period, namely prostacyclin and PGE₂. It is noteworthy that the urinary levels of
336 the main metabolite of PGE₂ were higher than for any of the other lipid mediators detected in
337 the urine.

338 The primary eicosanoids are potent biologically active mediators, however, they are
339 troublesome to measure since they are very rapidly metabolized and cleared from the
340 circulation (34). Also, following withdrawal of blood it has been shown that e.g. TXB₂ can be
341 generated *ex vivo* (29). This often makes the measurement of these primary compounds very
342 difficult, and the interpretation of such data ambiguous. Urine has emerged as a non-invasive
343 alternative and metabolism and urinary excretion of these compounds have been extensively
344 studied.(23, 38) The use of urinary excretion of eicosanoid metabolites is now well-
345 established.(10) As we used a mass spectrometry platform which has previously been

346 applied to study the urinary mediator excretion following allergen challenge (9), the current
347 results allow for comparison of differences and similarities with respect to the patterns of
348 excretion in response to these two different indirect triggers of bronchoconstriction.
349 Following allergen challenge the levels of CysLT, and metabolites of PGD₂ and TXB₂ were
350 all increased but there was no increase in levels of PGE₂ or its metabolites. For the
351 bronchoconstrictive mediators PGD₂ and CysLT, the results in this EVH study are thus
352 concordant with the findings in the allergen inhalation challenge study, whereas we could not
353 find significant increases of TXB₂ or its metabolites following EVH. The discrepancies in
354 mediator excretion following EVH and allergen challenge respectively, suggests differences
355 in the cells activated by the different challenges.

356 For PGE₂ we observed increases of both the primary mediator and its most abundant
357 metabolite tetranor-PGEM. This is distinctly different to what was found following allergen
358 challenge.(9) The finding of PGE₂ excretion following EVH but not allergen challenge might
359 be explained by the mechanisms of the challenges. The reactions to both of the challenges are
360 initiated by mast cell activation as evident by the uniform excretion of PGD₂, but whereas the
361 allergen challenge is an IgE-dependent specific mast cell activation, EVH activates mast cells
362 through changes in osmolarity.(2) The change in local tissue osmolarity is likely to induce
363 excretion of PGE₂ from other cells in the airways and in particular from airway epithelial
364 cells.(14, 16) In contrast, there was no increase in the excretion of PGF_{2α} which is consistent
365 with previous findings in plasma following exercise challenge (3), and, again underscores the
366 specificity of the pattern of excretion of lipid mediators.

367 The observation that pre-treatment with inhalation of PGE₂ has been shown to inhibit
368 the response to exercise challenge(25) as well as the inhibiting effect of NSAID pre-treatment
369 on the development of refractoriness(24, 26, 36), has led to the speculation PGE₂ is the key
370 mediator of refractoriness. However, there are no previous reports on increased excretion of

371 PGE₂ following exercise challenge in asthmatics, rather a decrease in the levels of PGE₂ has
372 been seen in induced sputum.(13) Increased levels have been seen in male subjects following
373 exercise in exhaled breath condensate (EBC) but the same was not seen in female
374 subjects.(31) As lipid mediators in EBC to a significant extent may reflect salivary
375 admixture, the data are inconclusive.(12)

376 In our study, the excretion of the abundant metabolite of PGE₂, tetranor-PGEM,
377 increased following challenge I but not following challenge II. This supports increased
378 excretion of PGE₂ from the lung following EVH challenge. Interestingly we also found that
379 primary PGE₂ increased in the urine following both of the challenges. The current concept is
380 that the kidney itself is exclusive source of urinary PGE₂. This view is based on previous
381 metabolic studies showing rapid metabolism of systemic PGE₂. Thus, primary PGE₂ was
382 only seen in the urine following renal artery infusion, but not after brachial vein infusion. (38)
383 However, the metabolic studies were done with relatively low doses of PGE₂ and it is likely
384 that the systemic load of PGE₂ following massive excretion from the airways during the EVH
385 challenge is much greater, explaining that a small proportion is excreted in the urine un-
386 metabolised.

387 Comparing the levels of urinary mediators between EIA and UPLC-MS/MS for LTE₄,
388 there were good correlations but absolute values were generally higher in EIA. For 11β-
389 PGF_{2α}, however, this metabolite was not at all detected in UPLC-MS/MS, but rather the
390 levels in EIA seemed to correspond to the levels of 2,3-dinor-PGF_{2α}, the metabolite to which
391 the antibody is cross-reactive. What is actually measured with the EIA for 11β-PGF_{2α}
392 therefore appears to be 2,3-dinor-PGF_{2α}. This has previously been noted in work from our
393 group,(27) and confirmed by others,(15) but the commonly used term has still been 11β-PGF_{2α}
394 because this is the name of the antibody in the commercially available kit. For the other
395 mediators analyzed both using EIA and UPLC-MS/MS there was in general a good agreement

396 between the methods as can be seen by similar patterns of excretion (figure 3) and good
397 correlations. In the Bland-Altman analysis(7) it was evident that with increasing
398 concentration the discrepancies between the EIA and UPLC-MS/MS results became larger.
399

400 We performed an initial study in order to optimize the conditions for mediator analysis
401 during the refractory period. From previous studies we know that refractoriness is greater the
402 sooner after the first challenge the second challenge is performed.(11) However, since there is
403 a lag between the excretion of mediators in the lung and the excretion in urine we needed to
404 extend the interval. Considering most mediators were back to baseline, and refractoriness was
405 still found, the 3 hours interval was sufficient. The subjects in study 2 displayed from no to
406 almost complete protection (2-70%) at the second challenge, which is in line with previous
407 findings that the degree of refractoriness is indeed a continuous response.(22)

408 Concerning the repeatability of the EVH challenge Bland-Altman analysis(7) revealed a
409 high variability between challenges, which is in line with findings by Price and
410 colleagues.(30) Considering that the between study day difference for several subjects were
411 >10%, the question arises about the usability of the EVH challenge as a predictive tool in the
412 diagnosis of EIB. The number of subjects with large differences was however smaller for the
413 6 minute protocol (Figure 6) which also caused a greater response. This makes the 6 min
414 challenge more suitable for diagnosis and drug intervention trials.

415 In conclusion, the consistent finding of increased levels of the bronchoconstrictive
416 mediators following two repeated eucapnic hyperventilation challenges makes decreased
417 mediator excretion following the second challenge an unlikely mechanism of refractoriness.
418 Our findings lend further support to the importance of PGE₂ and possibly also PGI₂ in the
419 development of refractoriness, and provide circumstantial support for our previous suggestion
420 of decreased responsiveness at the level of the airway muscle.(21, 22) The next step will need

421 to be specific interventions with e.g. subtype specific PGE₂ receptor (EP) agonists to define
422 the mechanisms in greater detail. We have recently found that low doses of PGE₂ via EP2
423 receptor activation has a long-lived inhibitory effect on mast cell dependent constriction of
424 human small airways.(33) A better understanding of the mechanism of this unique natural
425 protective mechanism may aid in the search for new treatment targets in asthma.

426

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435 **Disclosures** – none

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446 Figure legends:

447

448 **Figure 1.** Study design. EVH = eucapnic voluntary hyperpnea. U = urine sampling.

449

450 **Figure 2.** *Mediator excretion during study 1*

451 Levels of urinary mediators 11β -PGF_{2α} ng/mmol creatinine in urine during 1 hour (A) and 3

452 hour (B) protocol. LTE₄ ng/mmol creatinine during 1 hour (C) and 3 hour (D).

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454 **Figure 3.** *Extended analysis of the mediator excretion during study 2*

455 EIA and UPLC-MS/MS data. All values are presented as ng/mmol creatinine.

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472 **References:**

473 1. ATS/ERS recommendations for standardized procedures for the online and offline
474 measurement of exhaled lower respiratory nitric oxide and nasal nitric oxide, 2005. *Am J*
475 *Respir Crit Care Med* 171: 912-930, 2005.

476 2. **Anderson SD, and Daviskas E.** The mechanism of exercise-induced asthma is. *J*
477 *Allergy Clin Immunol* 106: 453-459, 2000.

478 3. **Anderson SD, Pojer R, Smith ID, and Temple D.** Exercise-related changes in plasma
479 levels of 15-keto-13,14-dihydro-prostaglandin F_{2α} and noradrenaline in asthmatic and
480 normal subjects. *Scand J Respir Dis* 57: 41-48, 1976.

- 481 4. **Anderson SD, Silverman M, Konig P, and Godfrey S.** Exercise-induced asthma. *Br J*
482 *Dis Chest* 69: 1-39, 1975.
- 483 5. **Argyros G, Roach J, Hurwitz K, Eliasson A, and Phillips Y.** Eucapnic voluntary
484 hyperventilation as a bronchoprovocation technique. *Chest* 109: 1520-1524, 1996.
- 485 6. **Balgoma D, Larsson J, Rokach J, Lawson JA, Daham K, Dahlen B, Dahlen SE,**
486 **and Wheelock CE.** Quantification of lipid mediator metabolites in human urine from asthma
487 patients by electrospray ionization mass spectrometry: controlling matrix effects. *Anal Chem*
488 85: 7866-7874, 2013.
- 489 7. **Bland M, and Altman D.** Statistical methods for assessing agreement between two
490 methods of clinical measurement. *Lancet* feb 8: 307-310, 1986.
- 491 8. **Brannan JD, Gulliksson M, Anderson SD, Chew N, and Kumlin M.** Evidence of
492 mast cell activation and leukotriene release after mannitol inhalation. *Eur Respir J* 22: 491-
493 496, 2003.
- 494 9. **Daham K, James A, Balgoma D, Kupczyk M, Billing B, Lindeberg A, Henriksson**
495 **E, FitzGerald GA, Wheelock CE, Dahlen SE, and Dahlen B.** Effects of selective COX-2
496 inhibition on allergen-induced bronchoconstriction and airway inflammation in asthma. *J*
497 *Allergy Clin Immunol* 134: 306-313, 2014.
- 498 10. **Dahlen SE, and Kumlin M.** Can asthma be studied in the urine? *Clin Exp Allergy* 28:
499 129-133, 1998.
- 500 11. **Edmunds AT, Tooley M, and Godfrey S.** The refractory period after exercise-induced
501 asthma: its duration and relation to the severity of exercise. *Am Rev Respir Dis* 117: 247-254,
502 1978.
- 503 12. **Gaber F, Acevedo F, Delin I, Sundblad BM, Palmberg L, Larsson K, Kumlin M,**
504 **and Dahlen SE.** Saliva is one likely source of leukotriene B4 in exhaled breath condensate.
505 *Eur Respir J* 28: 1229-1235, 2006.
- 506 13. **Hallstrand TS, Moody MW, Wurfel MM, Schwartz LB, Henderson WR, Jr., and**
507 **Aitken ML.** Inflammatory basis of exercise-induced bronchoconstriction. *Am J Respir Crit*
508 *Care Med* 172: 679-686, 2005.
- 509 14. **Harrington LS, Lucas R, McMaster SK, Moreno L, Scadding G, Warner TD, and**
510 **Mitchell JA.** COX-1, and not COX-2 activity, regulates airway function: relevance to aspirin-
511 sensitive asthma. *FASEB J* 22: 4005-4010, 2008.
- 512 15. **Higashi N, Mita H, Ono E, Fukutomi Y, Yamaguchi H, Kajiwara K, Tanimoto H,**
513 **Sekiya K, Akiyama K, and Taniguchi M.** Profile of eicosanoid generation in aspirin-
514 intolerant asthma and anaphylaxis assessed by new biomarkers. *J Allergy Clin Immunol* 125:
515 1084-1091 e1086, 2010.
- 516 16. **Hjoberg J, Folkerts G, van Gessel SB, Hogman M, Hedenstierna G, and Nijkamp**
517 **FP.** Hyperosmolarity-induced relaxation and prostaglandin release in guinea pig trachea in
518 vitro. *Eur J Pharmacol* 398: 303-307, 2000.
- 519 17. **Joos GF, O'Connor B, Anderson SD, Chung F, Cockcroft DW, Dahlen B, DiMaria**
520 **G, Foresi A, Hargreave FE, Holgate ST, Inman M, Lotvall J, Magnussen H, Polosa R,**
521 **Postma DS, and Riedler J.** Indirect airway challenges. *Eur Respir J* 21: 1050-1068, 2003.
- 522 18. **Kippelen P, Larsson J, Anderson SD, Brannan JD, Dahlen B, and Dahlen SE.**
523 Effect of sodium cromoglycate on mast cell mediators during hyperpnea in athletes. *Med Sci*
524 *Sports Exerc* 42: 1853-1860, 2010.
- 525 19. **Kippelen P, Larsson J, Anderson SD, Brannan JD, Delin I, Dahlen B, and Dahlen**
526 **SE.** Acute effects of beclomethasone on hyperpnea-induced bronchoconstriction. *Med Sci*
527 *Sports Exerc* 42: 273-280, 2010.
- 528 20. **Kumlin M, Stensvad F, Larsson L, Dahlen B, and Dahlen SE.** Validation and
529 application of a new simple strategy for measurements of urinary leukotriene E4 in humans.
530 *Clin Exp Allergy* 25: 467-479, 1995.

- 531 21. **Larsson J, Anderson SD, Dahlen SE, and Dahlen B.** Refractoriness to exercise
532 challenge: a review of the mechanisms old and new. *Immunol Allergy Clin North Am* 33: 329-
533 345, viii, 2013.
- 534 22. **Larsson J, Perry CP, Anderson SD, Brannan JD, Dahlen SE, and Dahlen B.** The
535 occurrence of refractoriness and mast cell mediator release following mannitol-induced
536 bronchoconstriction. *J Appl Physiol* 110: 1029-1035, 2011.
- 537 23. **Liston TE, and Roberts LJ, 2nd.** Metabolic fate of radiolabeled prostaglandin D2 in a
538 normal human male volunteer. *J Biol Chem* 260: 13172-13180, 1985.
- 539 24. **Manning PJ, Watson RM, and O'Byrne PM.** Exercise-induced refractoriness in
540 asthmatic subjects involves leukotriene and prostaglandin interdependent mechanisms. *Am*
541 *Rev Respir Dis* 148: 950-954, 1993.
- 542 25. **Melillo E, Woolley KL, Manning PJ, Watson RM, and O'Byrne PM.** Effect of
543 inhaled PGE2 on exercise-induced bronchoconstriction in asthmatic subjects. *Am J Respir*
544 *Crit Care Med* 149: 1138-1141, 1994.
- 545 26. **O'Byrne PM, and Jones GL.** The effect of indomethacin on exercise-induced
546 bronchoconstriction and refractoriness after exercise. *Am Rev Respir Dis* 134: 69-72, 1986.
- 547 27. **O'Sullivan S, Mueller MJ, Dahlen SE, and Kumlin M.** Analyses of prostaglandin D2
548 metabolites in urine: comparison between enzyme immunoassay and negative ion chemical
549 ionisation gas chromatography-mass spectrometry. *Prostaglandins Other Lipid Mediat* 57:
550 149-165, 1999.
- 551 28. **O'Sullivan S, Roquet A, Dahlen B, Larsen F, Eklund A, Kumlin M, O'Byrne PM,**
552 **and Dahlen SE.** Evidence for mast cell activation during exercise-induced
553 bronchoconstriction. *Eur Respir J* 12: 345-350, 1998.
- 554 29. **Patrono C, Ciabattoni G, Pugliese F, Pierucci A, Blair IA, and FitzGerald GA.**
555 Estimated rate of thromboxane secretion into the circulation of normal humans. *J Clin Invest*
556 77: 590-594, 1986.
- 557 30. **Price OJ, Ansley L, and Hull JH.** Diagnosing exercise-induced bronchoconstriction
558 with eucapnic voluntary hyperpnea: is one test enough? *J Allergy Clin Immunol Pract* 3: 243-
559 249, 2015.
- 560 31. **Pucsok JM, Gyore I, Argay K, Huszar E, Barat E, Pucsok J, and Horvath I.** Effect
561 of exercise on levels of cyclo-oxygenase mediators in exhaled breath condensate in elite
562 athletes. *J Sports Med Phys Fitness* 47: 223-227, 2007.
- 563 32. **Rosenthal R.** Simplified eucapnic voluntary hyperventilation challenge. *J Allergy Clin*
564 *Immunol* 73: 676-679, 1984.
- 565 33. **Safholm J, Manson ML, Bood J, Delin I, Orre AC, Bergman P, Al-Ameri M,**
566 **Dahlen SE, and Adner M.** Prostaglandin E inhibits mast cell-dependent bronchoconstriction
567 in human small airways through the E prostanoid subtype 2 receptor. *J Allergy Clin Immunol*
568 2015.
- 569 34. **Samuelsson B, Granstrom E, Green K, Hamberg M, and Hammarstrom S.**
570 Prostaglandins. *Annu Rev Biochem* 44: 669-695, 1975.
- 571 35. **Smith CM, Anderson SD, and Seale JP.** The duration of action of the combination of
572 fenoterol hydrobromide and ipratropium bromide in protecting against asthma provoked by
573 hyperpnea. *Chest* 94: 709-717, 1988.
- 574 36. **Wilson BA, Bar-Or O, and O'Byrne PM.** The effects of indomethacin on
575 refractoriness following exercise both with and without a bronchoconstrictor response. *Eur*
576 *Respir J* 7: 2174-2178, 1994.
- 577 37. **Zetterquist W, Pedroletti C, Lundberg JON, and Alving K.** Salivary contribution to
578 exhaled nitric oxide. *Eur Respir J* 13: 327-333, 1999.
- 579 38. **Zipser RD, and Martin K.** Urinary excretion of arterial blood prostaglandins and
580 thromboxanes in man. *Am J Physiol* 242: E171-177, 1982.

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Table 1. Screening and Study day results
Study I

Patient No.	Screening					Study day - 1 hour protocol			Study day - 3 hours protocol		
	Sex	Age	FEV ₁ (L)	FEV ₁ (% predicted)	EVH	EVH I	EVH II	% Protection	EVH I	EVH II	% Protection
1	F	35	3.9	111	-11.0	-14.5	-15.0	-3	-14.8	-15.4	-4
2	F	52	2.3	86	-12.0	-8.3*	-11.0	*	-10.1	-2.6	74
3	F	25	2.5	74	-15.5	-11.9	-17.4	-46	-15.4	-17.7	-15
4	F	28	3.3	94	-29.3	-39.7	-23.9	40	-32.5	-27.7	15
5	M	43	3.2	84	-12.9	-19.5	-18.5	5	-14.4	-10.8	25
6	F	34	3.3	72	-23.4	-17.0	-13.3	22	-38.7	-41.1	-6
7	M	28	3.4	90	-42.0	-26.7	-12.2	54	-20.4	-12.2	40
8	M	27	4.7	104	-16.8	-19.1	-18.8	2	-20.7	-19.5	6
9	M	44	3.5	70	-16.5	-21.5	-14.5	33	-18.0	-13.9	22
10	M	27	4.6	102	-28.7	-18.8	-15.3	19	-19.7	-24.0	-22
11	M	39	4.4	104	-21.6	-10.8	-11.2	-4	-19.0	-12.5	34
12	M	39	3.0	91	-17.8	-7.5*	-16.3	*	-28.0	-21.1	25
13	M	44	4.7	117	-12.9	-6.5*	-8.8	*	-11.1	-8.2	26
14	M	41	3.8	93	-13.7	-15.5	-9.1	41	-17.2	-5.5	68
15	M	47	3.6	93	-12.0	-5.9*	-12.7	*	-2.8*	-4.8	*
16	M	43	5.1	101	-35.5	-9.3*	-13.3	*	-12.1	-12.9	-7
Mean±SEM		37±2	3.7±0.2	93±3%	-20.1±2.3%	-19.6±2.4%	-15.4±1.2% (NS)	15±8%	-19.5±2.1%	-16.3±2.5% £	19±7%

Study 2

Patient No.	Screening					Study day		
	Sex	Age	FEV ₁ (L)	FEV ₁ (% predicted)	EVH	EVH I	EVH II	% Protection
17	M	25	5.2	103	-17.6	-16.7	-11.9	29
18	M	22	5.1	99	-27.1	-23.4	-18.1	23
19	M	24	3.8	82	-39.5	-41.9	-29.7	29
20	M	22	5.2	108	-55.8	-62.2	-45.5	27
21	M	24	4.9	101	-16.8	-19.7	-16.6	16
22	F	30	2.4	94	-26.6	-30.0	-29.4	2
23	F	47	3.0	96	-18.1	-18.2	-9.7	47
24	F	33	3.9	102	-18.7	-28.1	-8.4	70
Mean±SEM		28±3	4.0±0.4	98±3%	-27.5±4.9%	-30.0±5.4%	-21.2±4.5% \$	30±7%

The percentage change of baseline value. * = Subjects excluded from the refractoriness calculations (<10% fall from baseline following challenge I). M = Male. F = Female. FEV₁ = forced expiratory volume in 1 second. EVH I = the first EVH (eucapnic voluntary hyperpnea) challenge, EVH II = the second EVH challenge after one or three hours. EVH I vs EVH II (NS = non-significant; £ = p=0.0195; \$ = p=0.0076)

Table 2. UPLC-MS/MS ng/mmol creatinine

	Baseline 1	Peak 1	Baseline 2	Peak 2
PGD₂	Not detected			
11 β -PGF _{2α}	Not detected			
2,3-dinor-PGF _{2α}	60 \pm 14	88 \pm 18*	56 \pm 13	89 \pm 20 [£]
Tetranor-PGDM	202 \pm 30	234 \pm 23	190 \pm 21	240 \pm 20 [£]
PGE₁	Not detected			
13,14-dihydro-15-keto-PGE ₁	Not detected			
PGE₂	17 \pm 3	33 \pm 5*	19 \pm 4	25 \pm 4 [£]
13,14-dihydro-15-keto-PGE ₂	Not detected			
Tetranor-PGEM	3193 \pm 1015	3435 \pm 966*	3087 \pm 932	3957 \pm 1016
PGF_{2α}	107 \pm 18	161 \pm 41	154 \pm 43	164 \pm 34
13,14-dihydro-15-keto-PGF _{2α}	Not detected			
Tetranor-PGFM	Not detected			
TXB₂	14 \pm 6	15 \pm 5	13 \pm 5	13 \pm 3
11-dihydro-TXB ₂	8 \pm 2	9 \pm 2	7 \pm 2	9 \pm 1
2,3-dinor-TXB ₂	68 \pm 18	81 \pm 17	59 \pm 14	83 \pm 18
8-iso-PGF_{2α}	26 \pm 2	33 \pm 4	29 \pm 5	33 \pm 4
2,3-dinor-8-iso-PGF _{2α}	147 \pm 7	184 \pm 15	163 \pm 18	187 \pm 10 [£]
8,12-iPF _{2α} -IV	430 \pm 48	615 \pm 116	547 \pm 96	577 \pm 61
LTB₄; LTB₄ metabolites[§]	Not detected			
LTC₄; LTD₄	Not detected			
LTE₄	7 \pm 1	10 \pm 1	6 \pm 2	12 \pm 3 [£]
EXC₄; EXD₄; EXE₄	Not detected			

*= significantly different from 1st baseline (p<0.05)

£ = significantly different from 2nd baseline (p<0.05)

§ = 6-trans-LTB₄; 20-OH-LTB₄; 20-CO₂H-LTB₄

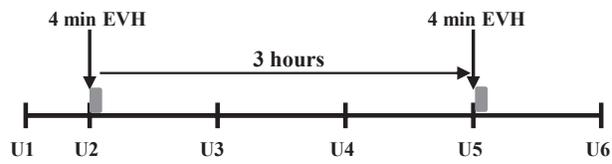
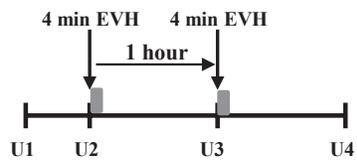
Table 3. Enzyme immunoassay ng/mmol creatinine

	Baseline 1	Peak 1	Baseline 2	Peak 2
PGD₂				
11 β -PGF _{2α}	65 \pm 15	80 \pm 16	58 \pm 15	71 \pm 14 [£]
Tetranor-PGDM	738 \pm 93	874 \pm 112*	815 \pm 98	880 \pm 103
PGE₂	49 \pm 7	77 \pm 9*	54 \pm 7	60 \pm 9
TXB₂	182 \pm 26	214 \pm 32	197 \pm 39	199 \pm 31
8-iso-PGF_{2α}	92 \pm 12	101 \pm 10	83 \pm 8	94 \pm 9 [£]
PGI₂				
6-keto-PGF _{1α}	158 \pm 23	210 \pm 38*	155 \pm 22	188 \pm 21 [£]
LTE₄	45 \pm 7	65 \pm 8*	53 \pm 10	57 \pm 7

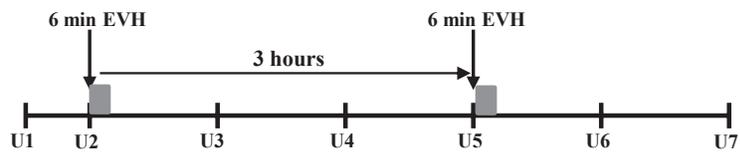
*= significantly different from 1st baseline (p<0.05)

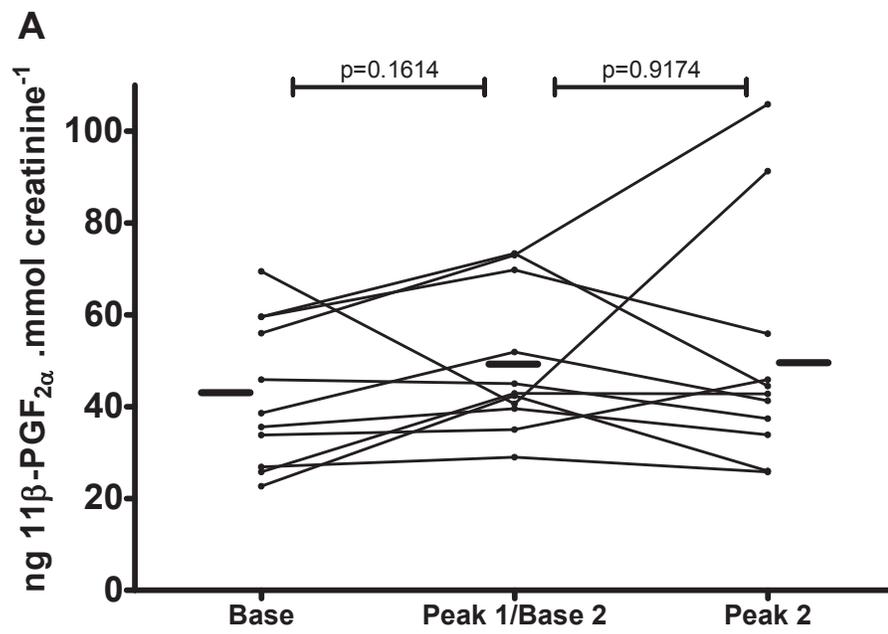
£ = significantly different from 2nd baseline (p<0.05)

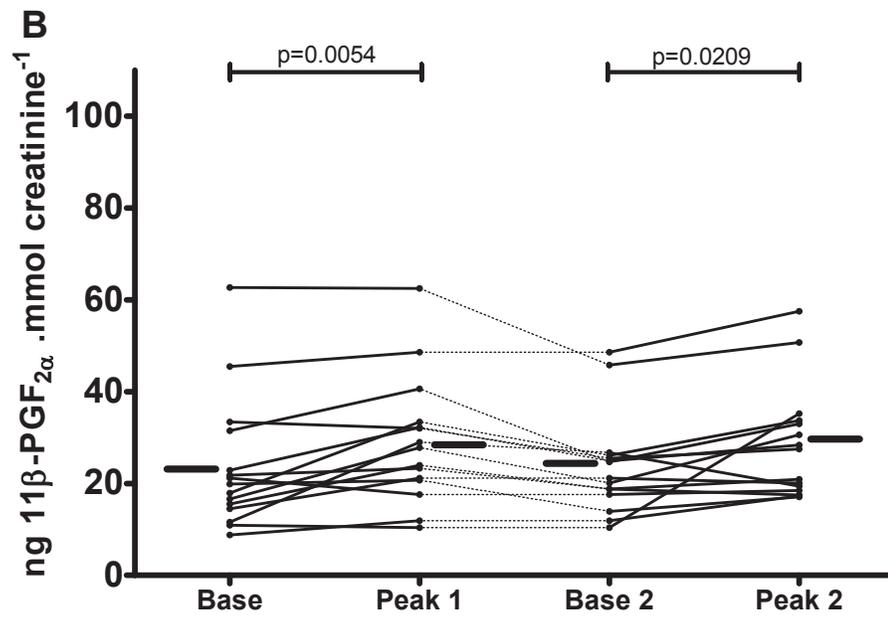
Study 1

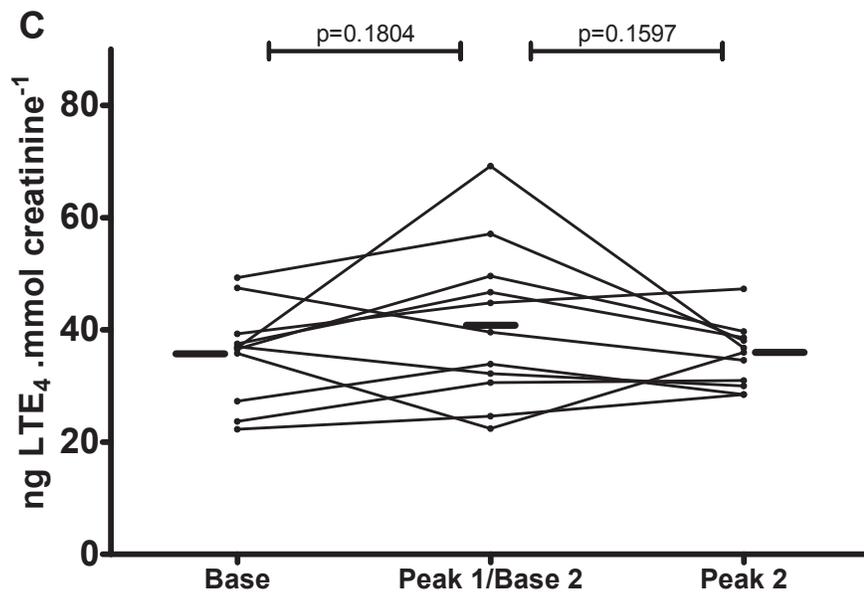


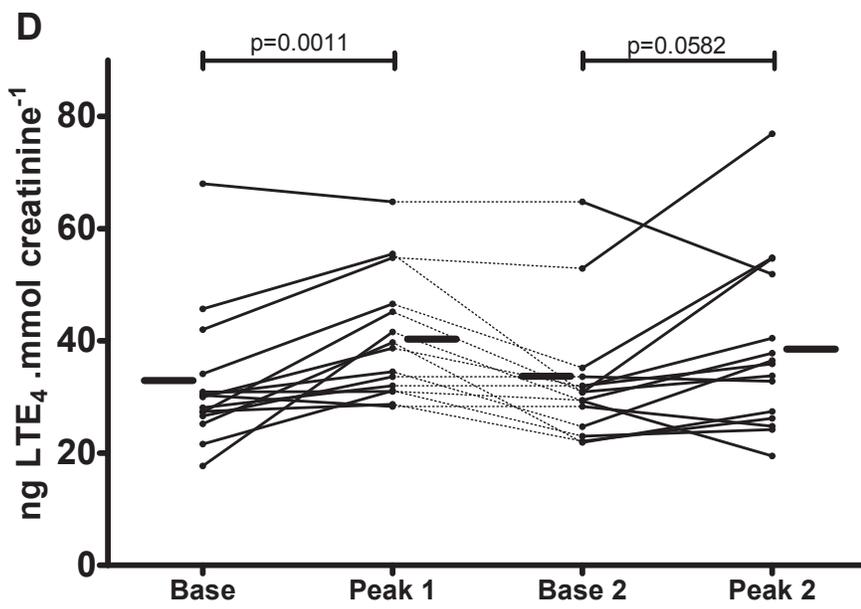
Study 2



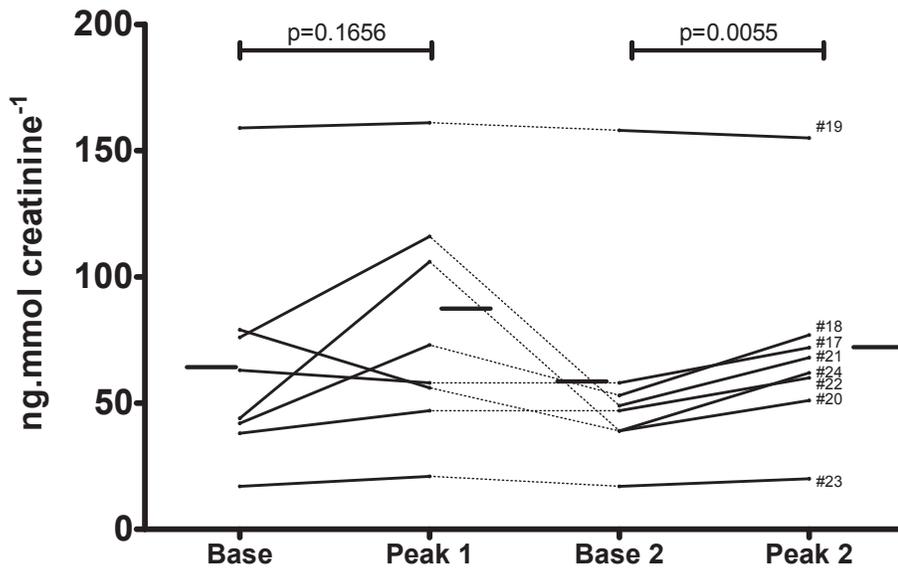




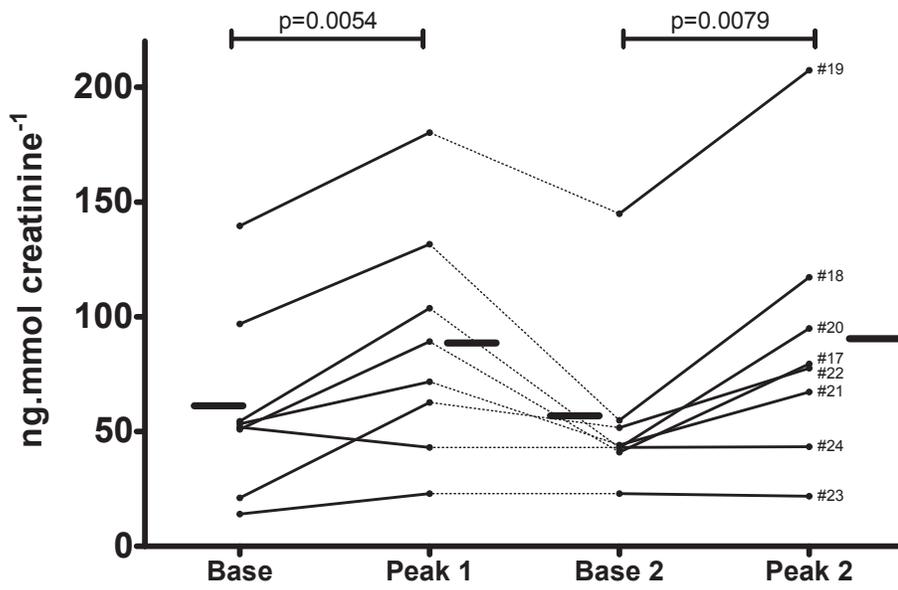


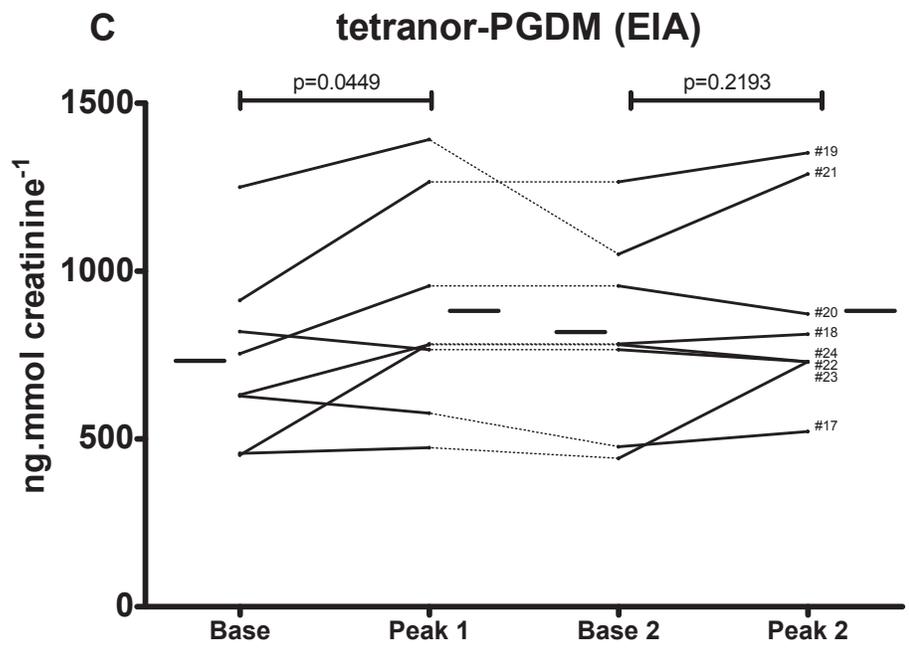


A 11b-PGF2a - EIA



B **2,3DN-PGF_{2α} (UPLC-MS/MS)**





D Tetranor-PGDM (UPLC-MS/MS)

